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Extended family: a caddisfly new to Saskatchewan, Canada with notes on the life history of *Neophylax splendens* (Trichoptera: Thremmatidae)

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Abstract—Streams draining the Cypress Hills support unique and understudied macroinvertebrate communities in Saskatchewan, Canada. Here, we report the discovery of a species of caddisfly new to the Cypress Hills and Saskatchewan, *Neophylax splendens* Denning (Trichoptera: Thremmatidae). Larvae were collected early in May 2012, and are found to enter pre-pupal diapause in mid-June until mid-September. Larvae were identified as *N. splendens* by morphological characters and verified with genetic analysis. Its occurrence strengthens the biogeographical link between the montane regions in British Columbia, Canada and Utah, United States of America with the southwest corner of Saskatchewan. This study highlights the importance of seasonal sampling, resolute species level identifications in biological surveys and the use of genetic analyses to obtain this level of identification.

Introduction

Species-level investigations are invaluable to aquatic ecology studies of community structure, including food web research, biodiversity surveys, and ecosystem health assessments (Rosenberg *et al.* 1986; Lenat and Resh 2001). Although the need for accurate species-level identifications is increasing in the field of aquatic entomology, the number of expert taxonomists is declining (New 1996; Stribling *et al.* 2003). Thus, accurate identifications using morphological characters alone often prove difficult to obtain due to lack of taxonomic expertise, significant variation in regional phenotypes and damaged specimens (Packer *et al.* 2009). Furthermore, aquatic studies often prove challenging because the commonly collected immature stages are difficult to identify to species as many are cryptic and taxonomic species level keys often do not exist and/or are designed for only the male or adult stage.

Cryptic species are indistinguishable morphologically, however, the use of morphological features in conjunction with DNA barcoding can lead to definitive identifications. (Pauls *et al.* 2010; Pramual *et al.* 2011). DNA barcoding (Hebert *et al.* 2003a) is the analysis of a standardised molecular identification system that provides

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¹Corresponding author (e-mail: brittney.hoemsen@usask.ca). Subject Editor: Justin Schmidt doi:10.4039/tce.2014.46 species-level identifications when morphological identification is difficult. Mitochondrial molecular markers are ideal for species level identification because mitochondrial DNA (mtDNA) has a faster mutation rate than nuclear DNA, often revealing genetic differences between species where many nuclear molecular markers would remain identical. The most commonly used molecular marker for DNA barcoding is a short (~650 base pairs) segment of the mitochondrial cytochrome C oxidase subunit 1 (COI) gene (Hebert et al. 2003b). The COI gene has been used to successfully identify aquatic invertebrates such as springtails (Collembola) (Hogg and Hebert 2004), mosquitoes (Kumar et al. 2007), Chironomidae (Diptera) (Pfenninger et al. 2007; Sinclair and Gresens 2008), black flies (Diptera: Simuliidae) (Rivera and Currie 2009), mayflies (Ephemeroptera) (Ball et al. 2005; Webb et al. 2012), and caddisflies (Trichoptera) (Parker 2000; Zhou et al. 2009, 2011).

Many areas in Saskatchewan, Canada are inhabited by rare organisms that provide great insight into the biodiversity and ecosystem health of the region (Phillips et al. 2008). The Cypress Hills in southwestern Saskatchewan, in particular, supports rich and unique biodiversity that remains understudied. The area is a flat-topped plateau acting as a montane refuge in the prairies for unique species (Phillips et al. 2013). Many of these species are deemed pre-glacial relicts (Hilton 1985; Campbell and Peck 1990), anomalies for the prairies (Shorthouse 1991), representative of Rocky Mountain fauna (Russell 1951; Bird 1962; Newsome and Dix 1968), or "Sonoran" in origin (Lehmkuhl 1980), originating from the Colorado system in Utah, United States of America. Focussed studies on such unique ecological systems and taxa, especially those including life history of aquatic insects, are highly valuable to our knowledge of the ecology of streams in the Northern Great Plains (Benke et al. 1984).

During a focussed ecological study of Pine Cree Creek in the Cypress Hills we discovered a new family (Thremmatidae) and species of Trichoptera for Saskatchewan, *Neophylax splendens* Denning. Here we describe our methods of combining morphological identification and genetic barcoding to identify *N. splendens*, and provide detailed observations about its life history.

Methods

Description of Pine Cree Creek

Pine Cree (49°36'32.1"N, 108°45'43.0"W) is a first order, spring-fed stream that flows through the Pine Cree Regional Park of the Cypress Hills (altitude = 1070 m; Fig. 1). The riparian zone includes mixed wood forest of white spruce (Picea glauca (Moench) Voss (Pinaceae)), lodgepole pine (Pinus contorta Douglas ex Loudon (Pinaceae)), trembling aspen (Populus tremuloides Michaux (Salicaceae)), and balsam poplar (Populus balsamifera Linnaeus (Salicaceae)). The creek has a mean annual specific conductivity of 638 µS $(\pm 38 \,\mu\text{S})$, mean pH of 7.02 (± 0.80) , mean summer temperature of 4.5 °C (May-September) with maximum temperature of 16.4 °C. Riparian health averages 89% and is deemed healthy following Prairie Conservation Action Plan (2008) riparian health assessments.

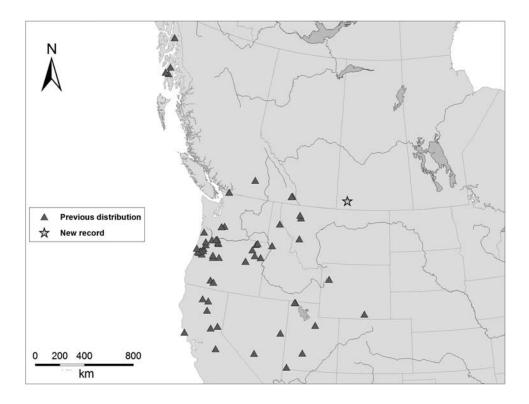
Sampling methods

We have conducted on-going annual fall sampling of Pine Creek and other streams in the Cypress Hills since 2006. A 500 µm mesh size D-frame net was used to sample riffle, run, and pool habitats. The site was visited monthly from May-October 2012, and three samples were collected in each riffle, run, and pool habitat using a 500 µm mesh size Hess sampler. In addition, an emergence trap was suspended over a productive pool habitat upstream of the Pine Cree Regional Park and emptied monthly. Samples were placed in 70% ethanol and sorted under 7× magnification. Specimens were identified using the keys of Clifford (1991), Vineyard (2005), and Merritt et al. (2008) and unique taxa were separated for further investigation. Diet was established through dissection of the gut and observation under 100× magnification.

Identification

Third instars (based on case length; Vineyard 2005) were collected in 5 May 2012. Specimens were sent to Michael Floyd at the United States Fish and Wildlife Service for verification and species identification. However, as the specimens were early instars with key characters including the setae on the submarginal row indiscernible, species-level identification was inconclusive (M. Floyd, United States Fish and Wildlife Service, personal communication). We therefore conducted genetic analysis

Fig. 1. Known distribution of *Neophylax splendens* (triangle markers) showing new record of the species from Pine Cree Creek, Saskatchewan, Canada (star marker). Records obtained through Barcode of Life Database (BOLD) (http://www.boldsystems.org) and Vineyard (2005).



to complement morphological characters for species determination (Flint and Kjer 2011). Specimens were selected for genetic analysis based on the following family and generic characters: long basal spurs on the tarsal claws, reduced metathorax Sa1 and sclerotised mesothoracic plate (Wiggins *et al.* 1985; Vineyard and Wiggins 1988; Vshivkova *et al.* 2007).

A single leg was removed from three selected specimens for DNA extraction and purification using DNeasy[®] Blood & Tissue Kit (Qiagen, Toronto, Ontario, Canada). Using the extracted DNA as a template, a 658 base pair region of the mitochondrial cytochrome oxidase I (COI) gene was targeted and amplified using two primers, IDT LCO1490 (5'-GGT CAA CAA ATC ATA AAG AAG ATA TTG G-3') and HCO2198 (5'-TAA ACT TCA GGG TGA CCA AAA AAT CA-3'). The polymerase chain reaction products were purified using ExoSAP-IT[®] procedure, and sequenced at the National Research Council Canada, Plant Biotechnology Institute, Saskatoon, Saskatchewan, Canada. The DNA sequences were

entered into the Barcode of Life Database (BOLD) (Ratnasingham and Hebert 2007) for taxonomic identification. All 191 *Neophylax* sequences in the BOLD database as of 15 January 2014 were downloaded as FASTA files and included in the analysis. Sequences were aligned using MUSCLE alignment (Edgar 2004) and TreeDyn (Dereeper *et al.* 2008, 2010) was used to generate a neighbourjoining tree.

Larval voucher specimens are deposited in collections at the Royal Saskatchewan Museum, Regina, Saskatchewan, Canada, and the Water Security Agency of Saskatchewan, Saskatoon, Saskatchewan, Canada.

Results and discussion

Larvae of *Neophylax splendens* were collected at Pine Cree Regional Park in the Cypress Hills, Saskatchewan, Canada 6 May 2012 (n = 3) and 6 June 2012 (n = 17) by I.D.P., B.M.H., and A.J.B. Thus, representing the first record of the family **Fig. 2.** Neighbour-joining tree depicting relationships between *Neophylax* specimens publicly available on Barcode of Life Database (BOLD) (BIN: BOLD:AAG9543) (http://www.boldsystems.org). Specimens recorded from Saskatchewan, Canada are new to the database. Species names are followed by their BOLD process ID number, the country and state in which they are recorded, and the BOLD BIN number. The scale bar indicates the length of a branch estimated to have experienced 0.03 substitutions per site.



Thremmatidae in Saskatchewan and extending the known distribution of this family over 500 km east of the Rocky Mountains (Fig. 1). Prepupal aggregations were observed 3 October 2012. No adult specimens were found in emergence traps nor through active searching in early October.

Morphological features identified the specimens as *Neophylax* species and COI sequences confirmed *N. splendens* (over 99.4% similarity) using the BOLD (http://www.boldsystem.org) with BIN: BOLD:AAG9543. Specimen and locality information are also available on GenBank (http://www.ncbi.nlm.nih.gov/genbank/) under the accession numbers KF367633–KF367635. Publicly available sequences within the same cluster as our specimens were used to generate a neighbour-joining tree (Fig. 2).

Life cycle

Neophylax splendens is typically found in small, first order streams at high elevations (Vineyard 2005), similar to Pine Cree Creek, Saskatchewan. All species of *Neophylax* are univoltine, hatching from eggs in early spring and emerging as adults in late fall (Vineyard 2005). In Pine Cree Creek, this caddisfly appears to have peak larval abundance in early June then enters pre-pupal diapause from late August until early October. A long diapause can allow larvae to avoid high summer temperatures, synchronise emergence (Corbet 1964; Masaki 1980), take advantage of increased food availability in fall (Beam and Wiggins 1987), or defer oviposition until the height of the drought period is over (Ross 1944). Pre-pupal aggregations were found on the undersides of rocks or debris in Pine Cree Creek in late September. These aggregations allow for avoidance of predation and parasitism (Otto and Svensson 1981) and can facilitate colonisation of other invertebrates (McCabe and Gotelli 2003). Pupation typically extends from two to six weeks with shorter emergence in cooler waters (Beam and Wiggins 1987). Adults generally emerge synchronously in early fall (Vineyard 2005). At Pine Cree Creek, no adults were collected in emergence traps, suggesting emergence was later than 3 October.

Diet

Larvae were collected in aggregations under woody debris and rocks in riffle habitat. Gut content analysis of *N. splendens* larvae from Pine Cree Creek revealed diatoms, periphyton, and fine organic particles of aufwuchs. Mandibles of *Neophylax* are simple scrapers with a brush of 10–15 long, slender setae along the mesal edge that serve to brush small particles into the mouth (Slack 1936). This supports their classification as obligate grazers (Sedell 1971; Cummins 1973).

Stream insect community

Many other Trichoptera were collected from Pine Cree Creek at the same time as N. splendens, including representatives of Glossosoma Curtis (Glossossomatidae), Rhyacophila Pictet (Rhyacophila), Hydropsyche Pictet (Hydropsychidae), Lepidostoma Rambur (Lepidostomatidae), Onocosmoecus Banks (Limnephilidae), Hesperophylax Banks (Limnephilidae), and Cheumatopsyche Wallengren (Hydropsychidae). These genera also have limited distributions in the northern and southern boreal region of Saskatchewan (Smith 1984). Neophylax rickeri and N. splendens are sister species (Vineyard 2005) and are typically found in sympatry but distributed along a longitudinal gradient, with N. splendens occurring upstream of N. rickeri (Vineyard 2005; Mendez and Resh 2008). However, we did not detect N. rickeri in our samples.

Distribution

Neophylax splendens is widely distributed in western North America, from southeastern Alaska to southern California and extending east in the Rocky Mountains into western Colorado and southcentral Wyoming (Fig. 1). It is the most common species of Neophylax in western North America but has not previously been recorded in the central part of the continent (Houghton et al. 2001). Much of the work on Trichoptera in Saskatchewan has focussed on the boreal streams and the North and South Saskatchewan Rivers (Smith 1975, 1984). No previous records of this species currently exist for Saskatchewan (Smith 1975); Royal Saskatchewan Museum collection; Water Security Agency of Saskatchewan voucher collection; AquaTax Consulting (Dale Parker, personal communication).

Neophylax splendens is typically found in cool, fast flowing streams (Vineyard 2005). The restricted locality of N. splendens in Saskatchewan could be due to the exceptional water quality of Pine Creek and unique characteristics of the Cypress Hills, including high elevation. Based on tolerance values for the northwest, N. splendens has a tolerance value of 3 (Barbour et al. 1999) indicating their sensitivity to human disturbance. Elevation could be the driver of the localised distribution of caddisflies (Williams 1991) in Pine Cree Creek, as it is at comparable elevations to the Utah, United States of America and British Columbia, Canada records (2040 and 1122 m, respectively). The occurrence of this species in the Cypress Hills strengthens the biogeographical link with montane regions such as Utah and British Columbia population. This supports Lehmkuhl's (1980) hypothesis of the insect fauna in the Cypress Hills being similar to montane regions such as Utah, forming the "Sonoran Connection".

Pine Cree Creek has been sampled each fall since 2006 as part of Saskatchewan's biomonitoring programme (Davies and Hanley 2010). Because this coincides with pre-pupal diapause or emergence time, this may explain why *N. splendens* has not been recorded to date. In the interests of expanding our knowledge of biodiversity in Saskatchewan, monitoring efforts should incorporate smaller order lotic systems. Currently, springs are understudied in Saskatchewan as sampling efforts focus largely on studying higher order systems due to their economical importance.

However, it is imperative to study the springs from which these rivers originate to better understand freshwater systems as a whole (Danks and Williams 1991). These smaller order streams can support rare and sensitive species, as we have shown with *N. splendens*. This species is adapted to cold springs and information relating to its life history and distribution is especially valuable in studies of biomonitoring and ecosystem health.

Conclusion

This study highlights the value of species-level identification in biological surveys and the use of genetic analysis in obtaining this taxonomic resolution. It also illustrates the inadequacy of annual, single season/date, sampling regimes to assess stream biodiversity.

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References

- Ball, S.L., Hebert, P.D.N., Burian, S.K., and Webb, J.M. 2005. Biological identifications of mayflies (Ephemeroptera) using DNA barcodes. Journal of the North American Benthological Society, 24: 508–524.
- Barbour, M.T., Gerritsen, J., Snyder, B.D., and Stribling, J.B. 1999. Rapid bioassessment protocols for use in streams and wadeable rivers: periphyton, benthic macroinvertebrates and fish, second edition [online]. EPA 841-B-99-002. United States Environmental Protection Agency, Office of Water, Washington, DC, United States of America. Available from http://water.epa.gov/scitech/monitoring/ rsl/bioassessment/index.cfm [accessed 8 July 2013].

- Beam, B.D. and Wiggins, G. 1987. A comparative study of the biology of five species of *Neophylax* (Trichoptera: Limnephilidae) in southern Ontario. Canadian Journal of Zoology, **65**: 1741–1754.
- Benke, A.C., Van Arsdall, T.C. Jr., Gillespie, D.M., and Parrish, F.K. 1984. Invertebrate productivity in a subtropical blackwater river: the importance of habitat and life history. Ecological Monographs, 54: 25–63.
- Bird, C. 1962. Bryophytes of the Cypress Hills Provincial Parks, Alberta and Saskatchewan. Canadian Journal of Botany, **40**: 573–587.
- Campbell, J. and Peck, S. 1990. *Omalonomus relictus*, an unusual new genus and new species (Coleoptera: Staphylinidae, Ohmalhnae) of blind rove beetle: a preglacial (Tertiary?) relict in the Cypress Hills, Alberta-Saskatchewan, Canada. The Canadian Entomologist, **122**: 949–961.
- Clifford, H.F. 1991. Aquatic invertebrates of Alberta. University of Alberta Press, Athabasca Hall, Edmonton, Alberta, Canada.
- Corbet, P.S. 1964. Temporal patterns of emergence in aquatic insects. The Canadian Entomologist, **96**: 264–279.
- Cummins, K.W. 1973. Trophic relations of aquatic insects. Annual Review of Entomology, 18: 183–206.
- Danks, H. and Williams, D.D. 1991. Arthropods of springs, with particular reference to Canada: synthesis and needs for research. Memoirs of the Entomological Society of Canada, **123**: 203–217.
- Davies, H. and Hanley, P.T. 2010. 2010 State of the watershed report. Saskatchewan Watershed Authority, Moose Jaw, Saskatchewan, Canada.
- Dereeper, A., Audic, S., Claverie, J.M., and Blanc, G. 2010. BLAST-EXPLORER helps you building datasets for phylogenetic analysis. BMC Evolutionary Biology, **10**: 8.
- Dereeper, A., Guignon, V., Blanc, G., Audic, S., Buffet, S., Chevenet, F., *et al.* 2008. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Research. Server issue: W465-9. Epub (PubMed). Available from http://www. phylogeny.fr/ [accessed 29 November 2013].
- Edgar, R.C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Research, **32**: 1792–1797.
- Flint, O.S. Jr and Kjer, K.M. 2011. A new species of *Neophylax* from Northern Virginia, USA (Trichoptera: Uenoidae). Proceedings of the Entomological Society of Washington, **113**: 7–13.
- Hebert, P.D.N., Cywinska, A., and Ball, S.L. 2003a. Biological identifications through DNA barcodes. Proceedings of the Royal Society of London. Series B: Biological Letters, **270**: 313–321.
- Hebert, P.D.N., Ratnasingham, S., and de Waard, J.R. 2003b. Barcoding animal life: cytochrome c oxidase subunit 1 divergences among closely related species. Proceedings of the Royal Society of London. Series B: Biological Letters, **270**: S96–S99.

- Hilton, D.F. 1985. Dragonflies (Odonata) of Cypress Hills Provincial Park, Alberta and their biogeographic significance. The Canadian Entomologist, 117: 1127–1136.
- Hogg, I.D. and Hebert, P.D.N. 2004. Biological identification of springtails (Hexapoda: Collembola) from the Canadian Arctic, using mitochondrial DNA barcodes. Canadian Journal of Zoology, 82: 749–754.
- Houghton, D.C., Holzenthal, R.W., Monson, M.P., and MacLean, D.B. 2001. Updated checklist of the Minnesota caddisflies (Tricoptera) with geographic affinities. Transactions of the American Entomological Society, **127**: 495–512.
- Kumar, N.P., Rajavel, A., Natarajan, R., and Jambulingam, P. 2007. DNA barcodes can distinguish species of Indian mosquitoes (Diptera: Culicidae). Journal of Medical Entomology, 44: 1–7.
- Lehmkuhl, D. 1980. Temporal and spatial changes in the Canadian insect fauna: patterns and explanation: the prairies. The Canadian Entomologist, **112**: 1145–1159.
- Lenat, D.R. and Resh, V.H. 2001. Taxonomy and stream ecology – the benefits of genus- and specieslevel identifications. Journal of the North American Benthological Society, **20**: 287–298.
- Masaki, S. 1980. Summer diapause. Annual Review of Entomology, 25: 1–25.
- McCabe, D.J. and Gotelli, N.J. 2003. Caddisfly diapause aggregations facilitate benthic invertebrate colonization. Journal of Animal Ecology, 72: 1015–1026.
- Mendez, P.K. and Resh, V.H. 2008. Life history of *Neophylax rickeri* (Trichoptera: Uenoidae) in two northern California streams. Annals of the Entomological Society of America, **101**: 573–584.
- Merritt, R.W., Cummins, K.W., and Berg, M.B. 2008. An introduction to the aquatic insects of North America. Kendall/Hunt, Dubuque, Indiana, United States of America.
- New, T. 1996. Taxonomic focus and quality control in insect surveys for biodiversity conservation. Australian Journal of Entomology, 35: 97–106.
- Newsome, R. and Dix, R.L. 1968. The forests of the Cypress Hills, Alberta and Saskatchewan, Canada. American Midland Naturalist, **80**: 118–185.
- Otto, C. and Svensson, B. 1981. Why do *Potamophylax cingulatus* (Steph.) (Trichoptera) larvae aggregate at pupation? Proceedings of the Third International Symposium on Trichoptera, **285**: 285–291.
- Packer, L., Gibbs, J., Sheffield, C., and Hanner, R. 2009. DNA barcoding and the mediocrity of morphology. Molecular Ecology Resources, 9: 42–50.
- Parker, C.R. 2000. *Neophylax kolodskii* (Trichoptera: Uenoidae), a new species from the Great Smoky Mountains National Park, USA. Aquatic Insects, 22: 271–274.
- Pauls, S.U., Blahnik, R.J., Zhou, X., Wardwell, C.T., and Holzenthal, R.W. 2010. DNA barcode data confirm new species and reveal cryptic diversity in Chilean Smicridea (*Smicridea*) (Trichoptera: Hydropsychidae). Journal of the North American Benthological Society, **29**: 1058–1074.

- Pfenninger, M., Nowak, C., Kley, C., Steinke, D., and Streit, B. 2007. Utility of DNA taxonomy and barcoding for the inference of larval community structure in morphologically cryptic *Chironomus* (Diptera) species. Molecular Ecology, **16**: 1957–1968.
- Phillips, I.D., Parker, D.W., Hoemsen, B.M., Bell, A.J., and Chivers, D.P. 2013. Biological notes and range expansion of the non-biting midge *Odontomesa fulva* (Kieffer) (Diptera: Chironomidae). Western North American Naturalist, **73**: 244–247.
- Phillips, I.D., Parker, D.W., and McMaster, G. 2008. The aquatic invertebrate fauna of a northern prairie river: range extensions and water quality characteristics. Western North American Naturalist, 68: 173–185.
- Prairie Conservation Action Plan. 2008. Riparian health assessment: streams and small rivers. Regina, Saskatchewan [online]. Available from http://www. pcap-sk.org/docs/5_resandlit/Streams_and_Small_ Rivers-Green.pdf [accessed 12 July 2013].
- Pramual, P., Wongpakam, K., and Adler, P.H. 2011. Cryptic biodiversity and phylogenetic relationships revealed by DNA barcoding of Oriental black flies in the subgenus *Gomphostilbia* (Diptera: Simuliidae). Genome, **54**: 1–9.
- Ratnasingham, S. and Hebert, P.D.N. 2007. BOLD: The barcode of life data system (http://www. barcodinglife.org). Molecular Ecology Notes, 7: 355–364.
- Rivera, J. and Currie, D.C. 2009. Identification of Nearctic black flies using DNA barcodes (Diptera: Simuliidae). Molecular Ecology Resources, 9: 224–236.
- Rosenberg, D.M., Danks, H., and Lehmkuhl, D.M. 1986. Importance of insects in environmental impact assessment. Environmental Management, 10: 773–783.
- Ross, H.H. 1944. The caddisflies, or Trichoptera of Illinois. Bulletin of Illinois Natural History Survey, 23: 1–326.
- Russell, L. 1951. Land snails of the Cypress Hills. Canadian Field Naturalist, **65**: 174–175.
- Sedell, J.R. 1971. Trophic ecology and natural history of *Neophylax concinnus* and *N. oligius* (Trichoptera: Limnephilidae). Ph.D. thesis. University of Pittsburgh, Pittsburgh, Pennsylvania, United States of America.
- Shorthouse, J. 1991. An unusual population of galls of *Diplolepis polita* (Hymenoptera: Cynipidae) in the Cypress Hills of southeastern Alberta. Canadian Field Naturalist, **105**: 542–549.
- Sinclair, C.S. and Gresens, S.E. 2008. Discrimination of *Cricotopus* species (Diptera: Chironomidae) by DNA barcoding. Bulletin of Entomological Research, **98**: 555–563.

- Slack, H. 1936. The food of caddisfly (Trichoptera) larvae. The Journal of Animal Ecology, **5**: 105–115.
- Smith, D.H. 1975. The taxonomy of the Trichoptera (caddisflies) of the Saskatchewan River in Saskatchewan. M.Sc. thesis. University of Saskatchewan, Saskatoon, Saskatchewan, Canada.
- Smith, D.H. 1984. Systematics of Saskatchewan Trichoptera larvae with emphasis on species from boreal streams. Ph.D. thesis. University of Saskatchewan, Saskatoon, Saskatchewan, Canada.
- Stribling, J.B., Moulton, S.R., and Lester, G. 2003. Determining the quality of taxonomic data. Journal of the North American Benthological Society, 22: 621–631.
- Vineyard, R.N. 2005. The caddisfly genus *Neophylax* (Trichoptera, Uenoidae). Royal Ontario Museum, Toronto, Ontario, Canada.
- Vineyard, R.N. and Wiggins, G.B. 1988. Further revision of the caddisfly family Uenoidae (Trichoptera): evidence for inclusion of Neophylacinae and Uenoidae. Systematic Entomology, **13**: 361–372.
- Vshivkova, T.S., Morse, J.C., and Ruiter, D. 2007. Phylogeny of Limnephilidae and composition of the genus *Limnephilus* (Limnephilidae: Limnephilinae, Limnehilini). *In* Proceedings of the XIIth international symposium on Trichoptera, 18–22 June 2006, Mexico City, Mexico. *Edited by* J. Bueno-Soria, R. Barba-Alvarez, and B. Armitage. The Caddis Press, Columbus, Ohio, United States of America. Pp. 309–319.
- Webb, J.M., Jacobus, L.M., Funk, D.H., Zhou, X., Kondratieff, B., Geraci, C.J., *et al.* 2012. A DNA barcode library for North American Ephemeroptera: progress and prospects. Public Library of Science One, 7: e38063.
- Wiggins, G.B., Weaver, J.S., and Unzicker, J.D. 1985. Revision of the caddisfly family Uenoidae (Trichoptera). The Canadian Entomologist, **117**: 763–800.
- Williams, N.E. 1991. Geographical and environmental patterns in caddisfly (Trichoptera) assemblages from coldwater springs in Canada. Memoirs of the Entomological Society of Canada, **123**: 107–124.
- Zhou, X., Adamowicz, S.J., Jacobus, L.M., DeWalt, R.E., and Hebert, P.D.N. 2009. Towards a comprehensive barcode library for Arctic life-Ephemeroptera, Plecoptera, and Trichoptera of Churchill, Manitoba, Canada. Frontiers in Zoology, 6: 1–30.
- Zhou, X., Robinson, J.L., Geraci, C.J., Parker, C.R., Flint, O.S. Jr, Etnier, D.A., *et al.* 2011. Accelerated construction of a regional DNA-barcode reference library: caddisflies (Trichoptera) in the Great Smoky Mountains National Park. Journal of the North American Benthological Society, **30**: 131–162.